

Original Article

Prevalence and Antimicrobial Susceptibility of Bacteria Isolated from Surgical Site and Bloodstream Infections of Hospitalized Patients at a Tertiary Heart Center

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ABSTRACT

Background and Objectives: Bacterial bloodstream infections (BSIs) and surgical site infections (SSIs) are among the most common nosocomial infections with high mortality and morbidity. We aimed to evaluate the frequency of various species among BSIs and SSIs at Tehran Heart Center, Tehran, Iran.

Methods: Patients with localized or systemic infections that became evident 48 hours or more after hospitalization were included. Data were prospectively collected in 4 intensive care units (ICUs), 5 cardiac care units (CCUs), 7 post-CCUs, and 5 surgical wards during two consecutive years in 2008 and 2009. Approximately 18414 coronary angiography and 7393 open-heart surgeries were done within this period. Antimicrobial susceptibility testing was performed by the Kirby–Bauer disk diffusion method, in accordance with the Clinical and Laboratory Standards Institute (CLSI) guidelines.

Results: Among 212 detected patients with SSI and/or BSI in the year 2008, 138 had hospital acquired infection (HAI) and 74 had non-HAI while these figures for 2009 was 165/270 and 105/270, respectively. *Staphylococcus aureus* (21.5%) and *Enterobacter* spp. (16.5%) were two most common pathogens responsible for hospital acquired BSIs while *S. aureus* (20.6%) and *S. epidermidis* (20.6%) were corresponding isolates responsible for community acquired BSIs. *Staphylococcus aureus* (53.3%) and *Escherichia coli* (11.0%) were the two most common pathogens responsible for hospital acquired SSIs in the year 2008, while *S. aureus* (49.0%) and *S. epidermidis* (11.0%) were the most frequently reported hospital acquired SSIs in 2009.

Conclusions: Making rational decisions about hospital infection control plans may reduce infection rates for bacteria with antimicrobial resistance.

Keywords: Prevalence, Nosocomial Infections, Surgical Wound Infection, Microbial Sensitivity Test

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Introduction

Bacterial bloodstream infections (BSIs) and surgical site infections (SSIs) are among the most common nosocomial infections and cause significant mortality, morbidity, and related socioeconomic costs worldwide (1-4). Nosocomial infections, also called “hospital-acquired infections (HAIs)”, are defined as infections not presenting or not incubating when the patient is hospitalized which are acquired during the hospital stay at least 48-72 hours following admission to health institution. The estimated annual BSIs rate in the United States is approximately 350,000 patients with associated mortality rate of 20% to 50% (2, 5). The magnitude of SSI in different parts of the world varies considerably with a reported incidence of 0.25% to 2.9% and mortality rate ranging from 10% to 29% (6); however, SSIs are the most common cause of nosocomial infection (7) and are reported to be a serious complication following cardiac surgery (8, 9) increasing the relevant intensive care unit (ICU) admission up to 60%, hospital readmission more than five times, and the probability of death twice as compared to similar patients without SSIs (10).

Various types of antibiotics are available. Excessive and inappropriate use of antibiotics in health care facilities and in the communities will expedite the development of antibiotic resistance. To formulate a useful approach to the problem of inappropriate drug use providing data about antibiotic use pattern is crucial (11). In Iran, the prevalence of prevalent bacterial pathogens and their antimicrobial resistance patterns has not been established by a valid nationwide surveillance study so far, and only some localized data from individual hospitals have been reported. In addition, most of the previous investigations in this regard are point prevalence studies though the gold standard for such studies is prospective, on-site continuous, hospital wide surveillance (12).

Therefore, we aimed to conduct a prospective,

on-site continuous, hospital wide study at Tehran Heart Center, a tertiary referral center, to evaluate the frequency of various species among surgical site and bloodstream infections. Our second purpose was to identify antimicrobial susceptibility pattern among the most prevalent isolates in order to provide a feasible guide for Iranian clinicians.

Materials and Methods

This study was conducted at Tehran Heart Center – a major referral and educational cardiac hospital affiliated to Tehran University of Medical Sciences. The center comprises four ICUs (86 beds), five cardiac care units (CCUs) (72 beds), seven post-CCUs (157 beds) and five surgical wards (120 beds). Data were collected prospectively for two consecutive years from 2008 to 2009. In the two years period, approximately 18414 coronary angiography and 7393 open-heart surgeries were performed in the hospital. All of our patients are routinely followed-up by our surgeons after the operation for one month if no implant is left in place and for one year if implant (prosthetic valves, nonhuman vascular grafts, mechanical heart, and implantable cardioverter device) is in place. Patients with localized or systemic infections that became evident 48 hours or more after hospitalization were included in this study according to the Center for Disease Control and prevention (CDC, Atlanta, Ga., USA) definitions for HAI (13).

We investigated the two most common HAI types: SSIs, and BSIs. Detection of the infection was based on clinical findings as well as laboratory data and other tests according to a modified center of disease control and prevention (CDC)’s criteria (Table 1). Required data were collected from the HAI patients as described previously (7). Briefly, data gathered by on-site observation of the Infection Control Team, by using hospital information system (HIS), by using laboratory information system (LIS) reports, and also by using the hospital surgery database.

Table 1- Prevalent microorganisms isolated from blood cultures in total and in hospital acquired infections subgroup separately

| Bacterial species or group | No. of isolates | Percent of isolates |
|-----------------------------------|-----------------|---------------------|
| Year 2008 | | |
| Hospital acquired | | |
| <i>Staphylococcus aureus</i> | 14 | 25.0 |
| <i>Enterobacter spp.</i> | 10 | 17.9 |
| <i>Enterococcus spp.</i> | 7 | 12.5 |
| <i>Staphylococcus epidermidis</i> | 6 | 10.7 |
| Community acquired | | |
| <i>Staphylococcus aureus</i> | 16 | 23.2 |
| <i>Staphylococcus epidermidis</i> | 11 | 15.9 |
| <i>Enterobacter spp.</i> | 10 | 14.5 |
| <i>Klebsiella spp.</i> | 8 | 11.6 |
| Year 2009 | | |
| Hospital acquired | | |
| <i>Staphylococcus aureus</i> | 12 | 18.5 |
| <i>Enterobacter spp.</i> | 10 | 15.4 |
| <i>Acinetobacter spp.</i> | 9 | 13.8 |
| <i>Pseudomonas aeruginosa</i> | 8 | 12.3 |
| Community acquired | | |
| <i>Staphylococcus epidermidis</i> | 22 | 24.2 |
| <i>Staphylococcus aureus</i> | 17 | 18.7 |
| <i>Acinetobacter spp.</i> | 10 | 11.0 |
| <i>Escherichia coli</i> | 7 | 7.7 |

Antimicrobial susceptibility testing was performed by the Kirby–Bauer disk diffusion method, in accordance with the Clinical and Laboratory Standards Institute (CLSI) guidelines (M02-A9). All culture samples referred to laboratory was qualified by quality control criteria and samples with colonization or contamination were excluded. Results are presented separately for each year of study (2008 and 2009) descriptively as raw numbers and percentages by the site of infection. For the

statistical analysis, the statistical software SPSS version 13.0 for windows (SPSS Inc., Chicago, IL) was used.

Result

In the 2-year study period, 482 patients with SSI and/or BSI were detected with a mean age of 57 years. Among 212 detected patients with SSI and/or BSI in the year 2008, 138 (65.10%) had HAI and 74 (34.90%) had none–HAI while

these figures for 2009 was 165/270 (61.11%) and 105/270 (38.89%), respectively.

Blood stream infections. Of 283 bacterial growth positive blood cultures, 126 (44.5%) occurred in 2008 and 157 (55.5%) in 2009. The proportion of HAI was 56 (44.4%) in 2008 as compared to 65 (41.4%) in 2009. Ward-wise distribution of bloodstream isolates at ICUs were 41.1% in the year 2008 and 53.8% in the next year. The majority of hospital acquired BSIs in the present study were caused by gram-positive organisms (31.4%). The four most common pathogens responsible for hospital acquired and community acquired infections subgroups are shown in Table 1.

In the study period, *S. aureus* (21.5%), *Enterobacter* spp. (16.5%), *Acinetobacter* spp. (13.8%) and *Pseudomonas aeruginosa* (12.3%) were the four most common pathogens responsible for hospital acquired BSIs while *S. aureus* (20.6%), *S. epidermidis* (20.6%), *E. spp.* (10.0%), *A. spp.* (7.5%) were corresponding isolates responsible for community acquired BSIs.

Antimicrobial susceptibility among species isolated from blood culture in patients with HAI is listed in Table 2. All *S. aureus* strains recovered from our blood cultures were susceptible to vancomycin; resistance to gentamicin, ciprofloxacin, and clindamycin was also relatively uncommon in HAIs – 25% (3/12), 25% (4/16) and 33.3% (7/21), respectively. A similar pattern of antimicrobial susceptibility was found among community acquired *S. aureus* infections. Methicillin Resistant *S. aureus* (MRSA) rate was 57.7% for HAIs and 42.2% for non-HAI infections. Ciprofloxacin 20/20 (100%) and meropenem (100%) and imipenem 18/19 (94.7%) were the 3 most effective antibiotics among hospital

acquired *Enterobacter* strains and 19/19 (100%) demonstrated resistance to cefalotin whereas community acquired *Enterobacter* isolates, demonstrated considerable susceptibility to most of the applied antibiotics (not shown in the table). Unfortunately, most of applied antibiotics did not show a desirable efficacy against *Acinetobacter* isolates in this study. The most effective antibiotic was imipenem 6/13 (46.2%) and ciprofloxacin 5/13 (38.5%) among hospital acquired strains while this pattern for infections acquired from community were 63.6% susceptibility to ciprofloxacin and 54.5% to imipenem.

Surgical site infections. Of 199 bacterial growth positive cultures cultivated from surgical site specimens, 86 (43.2%) occurred in 2008 and 113 (56.8%) in 2009. With regard to type of the infections, 95.3% of these isolates in the year 2008 were HAI in comparison with 88.5% in the year 2009. The distribution pattern of surgical site isolates at our hospital was 42.7% from ICU patients and 57.3% from non-ICU inpatients in the year 2008. In the following year, the corresponding proportions were 53 and 37 per cent. The majority of SSIs in the present study were caused by gram-positive organisms.

Staphylococci was the most prevalent group of organisms recovered from our surgical sites cultures in the study period (>50%). *Staphylococcus aureus* (53.3%) and *Escherichia coli* (11.0%) were the two most common pathogens responsible for hospital acquired SSIs in the year 2008, while *S. aureus* (49.0%) and *S. epidermidis* (11.0%) were the most frequently reported hospital acquired surgical site isolates in 2009 (Table 3).

Table 2- Antimicrobial susceptibility of species or groups isolated from blood cultures among hospital acquired infections in the study period

| Microorganisms | Antimicrobial | Total No. | Susceptible | Intermediate | Resistant |
|-----------------------------------|-----------------------------------|-----------|-------------|--------------|-----------------------|
| <i>Staphylococcus aureus</i> | | 26 | | | |
| | Ciprofloxacin | 16 | 12 (75.0) | 0 | 1 (6.3) |
| | Clindamycin | 21 | 14 (66.7) | 0 | 7 (33.3) |
| | Erythromycin | 26 | 16 (61.5) | 0 | 10 (38.5) |
| | Gentamicin | 12 | 9 (75.0) | 0 | 3 (25.0) |
| | Vancomycin | 26 | 26 (100.0) | 0 | 0 |
| | Cefoxitin | 26 | 11 (42.3) | 0 | 15 (57.7) <i>MRSA</i> |
| <i>Staphylococcus epidermidis</i> | | 12 | | | |
| | Clindamycin | 9 | 2 (22.2) | 1 (11.1) | 6 (66.7) |
| | Erythromycin | 10 | 1 (10.0) | 0 | 9 (90.0) |
| | Gentamicin | 7 | 6 (85.7) | 0 | 1 (14.3) |
| | Vancomycin | 12 | 12 (100.0) | 0 | 0 |
| | Cefoxitin | 12 | 2 (16.7) | 0 | 10 (83.3) |
| | Ciprofloxacin | 9 | 6 (66.7) | 0 | 3 (33.3) |
| <i>Acinetobacter spp.</i> | | 13 | | | |
| | Amikacin | 12 | 3 (25.0) | 2 (16.7) | 7 (58.3) |
| | Cefalothin | 9 | 1 (11.1) | 0 | 8 (88.9) |
| | Ciprofloxacin | 13 | 5 (38.5) | 0 | 8 (61.5) |
| | Ceftriaxone | 13 | 2 (15.4) | 0 | 11 (84.6) |
| | Gentamicin | 13 | 3 (23.1) | 1 (7.7) | 9 (69.2) |
| | Cefepime | 13 | 2 (15.4) | 0 | 11 (84.6) |
| | ceftazidime | 13 | 0 | 2 (15.4) | 11 (84.6) |
| | imipenem | 13 | 6 (46.2) | 0 | 7 (53.8) |
| | Meropenem | 5 | 1 (20.0) | 0 | 4 (80.0) |
| | Ampicillin | 9 | 0 | 0 | 9 (100.0) |
| | Amoxicillin/ clavulanate | 8 | 1 (12.5) | 0 | 7 (87.5) |
| | Trimethoprim- sulfamethoxazole | 11 | 3 (27.3) | 0 | 8 (72.7) |
| | tobramycin | 6 | 1 (16.7) | 1 (16.7) | 4 (66.7) |
| <i>Enterobacter spp.</i> | | 20 | | | |
| | Cefalothin | 19 | 0 | 0 | 19 (100.0) |
| | Ciprofloxacin | 20 | 20 (100.0) | 0 | 0 |
| | Ceftriaxone | 17 | 12 (70.6) | 1 (5.9) | 4 (23.5) |
| | Gentamicin | 19 | 14 (73.7) | 0 | 5 (26.3) |
| | ceftazidime | 18 | 10 (55.6) | 4 (22.2) | 4 (22.2) |
| | Cefepime | 19 | 12 (63.2) | 1 (5.3) | 6 (31.6) |
| | imipenem | 19 | 18 (94.7) | 1 (5.3) | 0 |
| | Amikacin | 20 | 16 (80.0) | 2 (10.0) | 2 (10.0) |
| | Meropenem | 7 | 7 (100.0) | 0 | 0 |
| | Amikacin | 6 | 1 (16.7) | 0 | 5 (83.3) |
| | Ciprofloxacin | 7 | 2 (28.6) | 1 (14.3) | 4 (57.1) |

Table 3- Prevalent microorganisms isolated from surgical site in total and in hospital acquired infections subgroup separately

| Bacterial species or group | No. of isolates | Percent of isolates |
|-----------------------------------|-----------------|---------------------|
| Year 2008 | 82 | |
| Hospital acquired | | |
| <i>Staphylococcus aureus</i> | 48 | 58.5 |
| <i>Escherichia coli</i> | 13 | 15.9 |
| <i>Staphylococcus epidermidis</i> | 7 | 8.5 |
| <i>Pseudomonas aeruginosa</i> | 4 | 4.9 |
| Community acquired | 4 | |
| <i>Staphylococcus aureus</i> | 1 | 25.0 |
| <i>Escherichia coli</i> | 1 | 25.0 |
| <i>Staphylococcus epidermidis</i> | 1 | 25.0 |
| <i>Escherichia coli</i> inactive | 1 | 25.0 |
| Year 2009 | | |
| Hospital acquired | 100 | |
| <i>Staphylococcus aureus</i> | 49 | 49.0 |
| <i>Staphylococcus epidermidis</i> | 11 | 11.0 |
| <i>Escherichia coli</i> | 7 | 7.0 |
| <i>Pseudomonas aeruginosa</i> | 6 | 6.0 |
| <i>Acinetobacter</i> spp. | 6 | 6.0 |
| Community acquired | 13 | |
| <i>Staphylococcus aureus</i> | 11 | 84.6 |
| <i>Acinetobacter</i> spp. | 1 | 7.7 |
| <i>Klebsiella</i> spp. | 1 | 7.7 |

Susceptibility rates of antimicrobial agents applied for the most prevalent isolated bacterial strains are summarized in Table 4. Regarding hospital acquired *S. aureus* strains, susceptibility to vancomycin was (100.0%) by performing minimum inhibitory concentration (MIC) method and followed by susceptibility to gentamycin (78.6), clindamycin (77.4%), ciprofloxacin (72.0%) and erythromycin (70.5%). The rate of MRSA among hospital acquired SSIs was 56.5% in the current study. The antimicrobial agents' response against community acquired *S. aureus* strains was quite different with 88.9% rate of

MRSA was in this group of microorganisms. Imipenem revealed 100.0% effectiveness against *Escherichia coli* isolates but more than 70% resistance was found with cefalothin, ciprofloxacin, ceftazidime, and ceftriaxone. Notably, susceptibility rate of vancomycin for *S. epidermidis* was near 100% while this microorganism revealed about 60% resistance to erythromycin. Furthermore the rate of MRSE was 64.3% among our hospital acquired SSIs. Other antimicrobial-resistant pathogens encountered among our patients include *P. aeruginosa* resistant to cefepime (4/10; 40%).

Table 4- Antimicrobial susceptibility of species or groups isolated from surgical site among hospital acquired infections in two years

| Microorganisms | Antimicrobial | Total No. | Susceptible | Intermediate | Resistant |
|-----------------------------------|---------------|-----------|-------------|--------------|-----------------------|
| <i>Staphylococcus aureus</i> | | 97 | | | |
| | Cefoxitin | 85 | 37 (43.5) | 0 | 48 (56.5) <i>MRSA</i> |
| | Ciprofloxacin | 50 | 36 (72.0) | 6 (12.0) | 8 (16.0) |
| | Clindamycin | 84 | 65 (77.4) | 1 (1.2) | 18 (21.4) |
| | Erythromycin | 95 | 67 (70.5) | 2 (2.1) | 26 (27.4) |
| | Gentamicin | 42 | 33 (78.6) | 12 (2.4) | 8 (19.0) |
| | Vancomycin | 97 | 96 (99.0) | 0 | 1 (1.0) |
| <i>Escherichia coli</i> | | 20 | | | |
| | Amikacin | 20 | 14 (70.0) | 2 (10.0) | 4 (20.0) |
| | Cefalothin | 20 | 0 | 2 (10.0) | 18 (90.0) |
| | Ciprofloxacin | 20 | 3 (15.0) | 0 | 17 (85.0) |
| | Ceftriaxone | 19 | 3 (15.8) | 1 (5.3) | 15 (78.9) |
| | Gentamicin | 20 | 11 (55.0) | 0 | 9 (45.0) |
| | Cefepime | 19 | 6 (31.6) | 1 (5.3) | 12 (63.2) |
| | Ceftazidime | 20 | 4 (20.0) | 0 | 16 (80.0) |
| | Imipenem | 20 | 20 (100.0) | 0 | 0 |
| <i>Staphylococcus epidermidis</i> | | 18 | | | |
| | Vancomycin | 18 | 17 (94.4) | 0 | 1 (5.6) |
| | Erythromycin | 18 | 7(38.9) | 2 (11.1) | 9 (50.0) |
| | Cefoxitin | 14 | 5 (35.7) | 0 | 9 (64.3) <i>MRSE</i> |
| | Clindamycin | 15 | 10 (66.7) | 0 | 5 (33.3) |
| <i>Pseudomonas aeruginosa</i> | | 10 | | | |
| | Amikacin | 9 | 8 (88.9) | 0 | 1 (11.1) |
| | Tobramycin | 10 | 9 (90.0) | 0 | 1 (10.0) |
| | Imipenem | 10 | 8 (80.0) | 1 (10.0) | 1 (10.0) |
| | Ciprofloxacin | 9 | 8 (88.9) | 0 | 1 (11.1) |
| | Gentamicin | 10 | 9 (90.0) | 0 | 1 (10.0) |
| | Cefepime | 10 | 3 (30.0) | 3 (30.0) | 4 (40.0) |
| | Ceftazidime | 8 | 2 (25.0) | 4 (50.0) | 2 (25.0) |

Discussion

A number of factors contribute to the emergence of antimicrobial resistance in various hospital wards including the severity of patient illness, predisposition to nosocomial infections, cross-transmission of pathogens characteristic of critical care areas within the hospital, compromised membrane and skin barriers following the use of invasive devices (14). These factors extend length of hospital stay, and the widespread use

of prophylactic and therapeutic anti-infective agents. Surveillance of bacterial antimicrobial resistance is the most important challenges of our attempts to understand the dynamics of decreasing susceptibility to antibiotics in both hospital and community-acquired bacterial pathogens (15). Since there are a few data regarding bacterial resistance pattern in Iran, it is essential to prospectively evaluate the distribution of bacterial species isolated and their susceptibility pattern (16). In the current study, *S. aureus* and *Entro-*

bacter spp. were the two most common hospital acquired blood culture isolates in our hospital laboratory in the study period. The majority of hospital acquired BSIs were caused by gram-positive organisms.

American and European investigations have generally demonstrated *S. aureus* and *E. coli* as the two most common hospital acquired blood culture isolates (15, 17, 18) while another study in United States introduced coagulase-negative staphylococci as the most prevalent organism responsible for BSI with the overall 78.1% frequency of gram-positive bacteria (19). In an Iranian children medical center, Coagulase-negative staphylococci and *S. aureus* were prevalent microorganisms isolated from blood cultures examined while 72% of isolations were gram-positive bacteria (16). Coagulase-negative *Staphylococci* accounted for most isolated bacteria (24.61%), in both genders. The second most frequent isolated bacteria in adults were *S. aureus*, and in children was *Klebsiella pneumonia* (20).

With regard to susceptibility of gram-positive organisms recovered from our blood cultures, we found all *S. aureus* isolates remained fully to be susceptible to vancomycin (100%), showing that isolates with reduced susceptibility still remain rare local events. This finding is compatible with the results of European SENTRY study, the surveillance program conducted in the USA, Canada and Latin America by the same network, and to the results from other regions of the world (15, 17, 18, 21).

Among *S. aureus* isolates, the incidence of resistance to methicillin was 57.7% in our study similar to that observed by the SENTRY study in the Asia-Pacific region (17). A report from another city in Iran has also documented similar rates of methicillin resistance (22). Across Europe, methicillin resistance in *S. aureus* averaged 24% but with considerable regional variation from less than 5% to more than 50 % (15, 18). *Enterobacter* species showed the lowest

level of resistance to quinolones, meropenem and imipenem which is in line with the findings of other local and international studies (16-18).

In the present investigation, coagulase-negative staphylococci and *E. coli* were the most common pathogens isolated from SSIs accounting for more than half of the cases of hospital acquired postoperative infections in the study period. *Staphylococcus aureus* is a major pathogen in mediastinitis which is associated with preoperative contamination (23, 24).

Among *S. aureus* isolated from SSIs in our study, the number of MRSA was 56.5% while the rate of methicillin resistance for *S. epidermidis* was 64.3%. These results suggest a better condition in our hospital than that of other hospitals in this geographic region (14, 25). Practicing good hand hygiene is the cornerstone of any infection prevention and control programs, which plays an essential role in reducing the transmission of organisms. Contact precautions, environmental, equipment cleaning, and decontamination present a great risk for contamination with MRSA (26). All of the *S. aureus* and *Staphylococcus epidermidis* strains in our study had 100% susceptibility to vancomycin. This pattern is the same as findings of other Iranian studies (25, 27, 28).

Conclusion

Considerable differences in antimicrobial resistance do exist for each single hospital that reflects both antimicrobial usage pattern and infection control strategies. Making rational decisions about hospital infection control plans will reduce rates of infection with antimicrobial-resistant bacteria.

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