

Evaluation of the Relationship Between Expression of Villin and Gelsolin Genes and Axillary Lymph Node Metastasis in Patients with Breast Cancer

Armin Borhan^{1*}, Zohreh Nozarian¹, Alireza Abdollahi¹, Reza Shahsiah¹,
Hadiseh Mohammadpour², Arash Borhan²

1. Department of Pathology, Cancer Institute of Iran, Imam Khomeini Hospital, Tehran University of Medical Sciences, Tehran, Iran
2. Iran National Tumor Bank, Cancer Institute of Iran, Tehran University of Medical Sciences, Tehran, Iran

KEYWORDS

Breast cancer,
Gelsolin,
Metastasis,
Villin

Scan to discover online



Main Subjects:
Breast Pathology

Received 12 Feb 2020;

Accepted 12 April 2020;

Published Online 10 Oct 2020;

doi 10.30699/ijp.2020.121532.2322

ABSTRACT

Background & Objective: Nowadays, actin-binding proteins such as Villin and Gelsolin have been considered to be associated with aggressive tumors. This study mainly aims to determine the relationship between Gelsolin and Villin genes expression and metastasis of axillary lymph nodes in patients with breast cancer.

Methods: The included population consisted of 40 confirmed cases of female breast cancer (including 20 patients with breast cancer along with axillary lymph node metastasis and 20 patients without axillary lymph node metastasis). Expression of Villin and Gelsolin genes was evaluated using Real-time PCR and pre-designed primers.

Results: The mean expression level of Villin in groups with and without axillary lymph node metastasis was 3.33 ± 1.35 and 0.87 ± 0.88 , respectively ($P < 0.001$). The mean Gelsolin expression levels in both groups (with and without axillary lymph node metastasis) were 4.13 ± 2.40 and 1.00 ± 0.35 , respectively ($P < 0.001$). The significant relationships were independent of individuals' age.

Conclusion: Patients with axillary lymph node metastasis may express significant higher level of Villin and Gelsolin genes.

Corresponding Information:

Armin Borhan, Department of Pathology, Cancer Institute of Iran, Imam Khomeini Hospital Complex, Keshavarz Blvd., Tehran, Iran. Tel: +98-2161192337, E-mail: drarmin.borhan@gmail.com

Copyright © 2021. This is an open-access article distributed under the terms of the Creative Commons Attribution- 4.0 International License which permits share, copy and redistribution of the material in any medium or format or adapt, remix, transform, and build upon the material for any purpose, even commercially.

Introduction

Breast cancer is generally considered as the most common malignancy among the female population, affecting 1.7 million women worldwide every year (1, 2) constituting one-fourth of cancers in women and 15% of all cancer-related deaths (3). One in 8 women will develop breast cancer during life (4). In developing countries such as Iran, breast cancer rate increases by about 5% every year (5). Breast cancer onset happens 10 years earlier in the Iranian female population (24 in 100000) (6, 7). Recently, the survival rate of breast cancer patients has significantly increased due to early detection of malignancy and multiple effective treatments including surgical approaches as well as various chemoradiotherapy agents (1, 8, 9). Villin, a 92.5 kDa protein, is a crucial member of actin-binding proteins generally found in the microvilli of brush borders of epithelial linings of different gastrointestinal organs such as stomach, pancreas gland, bile ducts and intestines; as well as corresponding adenocarcinomas (10, 11). It has been suggested that the expression of Villin gene is closely related to a higher rate of malignant tumors metastatic

progression, aggressive behavior of cancer and consequently poorer prognosis (12).

Gelsolin, a member of Gelsolin protein super family, is an 82 kDa protein likely expressed in various cell types containing six homologous subdomains, ultra-structurally. Gelsolin is found in intracellular and extracellular regions (13, 14). Variations in Gelsolin gene expression leads to cytoskeletal changes during cell differentiation and carcinogenesis (13, 15). Some of the previously conducted studies suggest the role of inhibiting Gelsolin gene in tumor suppression. Declined expression of the Gelsolin gene was related to invasive breast cancer in mentioned studies (16, 17).

As there are little studies regarding the role of Villin gene and Gelsolin gene in patients with breast cancer in the Iranian female population, we designed this study to evaluate the relationship between gene expression of Gelsolin and Villin and axillary lymph node metastasis in patients with breast cancer.

Materials and Methods

This cross-sectional descriptive-analytic study was conducted in Imam Khomeini hospital complex (affiliated hospital of Tehran University of medical sciences) between January 2017 and January 2018.

The inclusion criteria were: invasive ductal carcinoma of breast, grade 2 or 3.

The exclusion criteria were: previous chemotherapy and/or radiotherapy.

Informed consent forms were obtained from included cases.

A portion of each tissue sample was fixed in 10% formalin and subsequently blocked in paraffin. Hematoxylin and eosin stained slides were prepared for histopathological evaluation including tumor histology (type and grade) and presence or absence of axillary lymph node metastasis. It is necessary to mention that the tissue diagnosis of breast malignancy, as well as tumor histologic grade, was performed by histopathological evaluation, utilizing Bloom-Richardson grading system (18).

The other portion of tissue sample was freshly received and stored in liquid nitrogen for RNA extraction. To determine the level of mRNA of each of the Villin and Gelsolin indices, the total RNA from the

mentioned frozen tissue sample was extracted using the Invitrogen TRIzol kit (Thermo Fisher Scientific, United States), as stated by the following procedure. Tissue samples were homogenized and vortex mixed for 10 seconds followed by addition of 200 μ L of Chloroform. Samples were centrifuged for 15 minutes at 1200 rpm and 4°C. The supernatant component was gathered and centrifuged after adding 500 μ L of isopropanol. The RNA pellets were washed using Ethanol and air dried. Subsequently, the RNA pellets were eluted with Diethyl Pyrocarbonate (DEPC) treated water. Afterwards, extracted RNA was quantified using a NanoDrop ND-2000 spectrophotometer (NanoDrop Technologies, Wilmington, DE). Quality validation of the extracted RNA was also done by gel electrophoresis technique. Using the RevertAid First Strand cDNA synthesis kit (Thermo Fisher Scientific, United States), complementary DNA (cDNA) was generated from mRNA. Mentioned synthesized cDNA was evaluated with housekeeping gene control. To monitor gene expression of two indices of the study (Villin gene and Gelsolin gene), real-time PCR (Takara Bio Inc., Japan) was performed using pre-designed primers (Table 1). Finally, values were determined from two indices in two groups of study.

Table 1. Pre-designed primer sequences used to determine genes expression (19)

Gene name	Primer sequence
Villin	F: 5'-GGCCAGCCAAGATGAAATTA-3'
	R: 5'-CTCAAAGGCCTTGGTGTGT-3'
Gelsolin	F: 5'-AGGTGGAGGAAGGCAGTGAAC-3'
	R: 5'-CGAAGAGGCGAGGAGGATGAG-3'

We applied SPSS software (version 19.0, IBM Corp., Armonk, NY, USA) for statistical data analysis. Mann-Whitney U test was also applied to contrast continuous variables. Statistical significance was set at P-value<0.05.

Results

The population involved in this study included 40 women with histologically confirmed breast cancer and mean age of 59.4 \pm 10.6 (36-78) years. Half of the

patients were also dealing with axillary lymph node involvement. There was a significant difference regarding Villin and Gelsolin gene expressions in patients with and without axillary lymph metastasis (Table 2) (Figures 1 and 2).

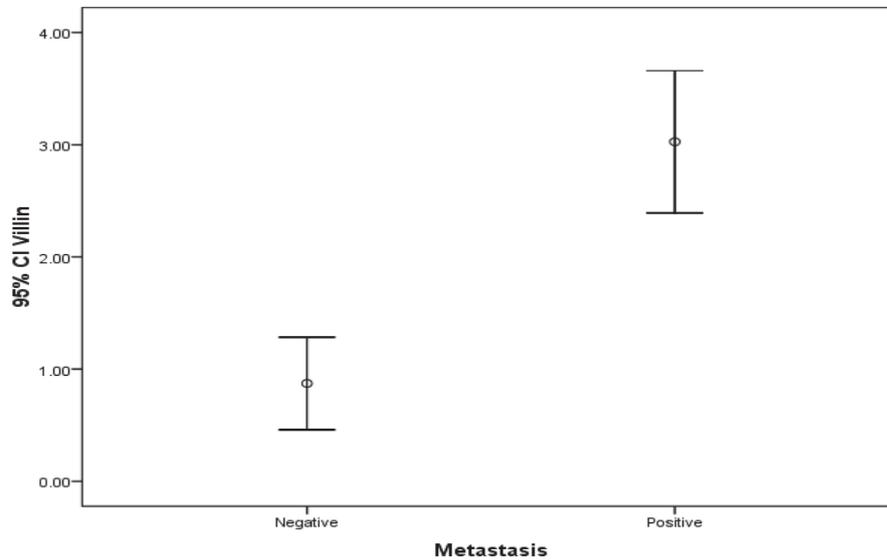
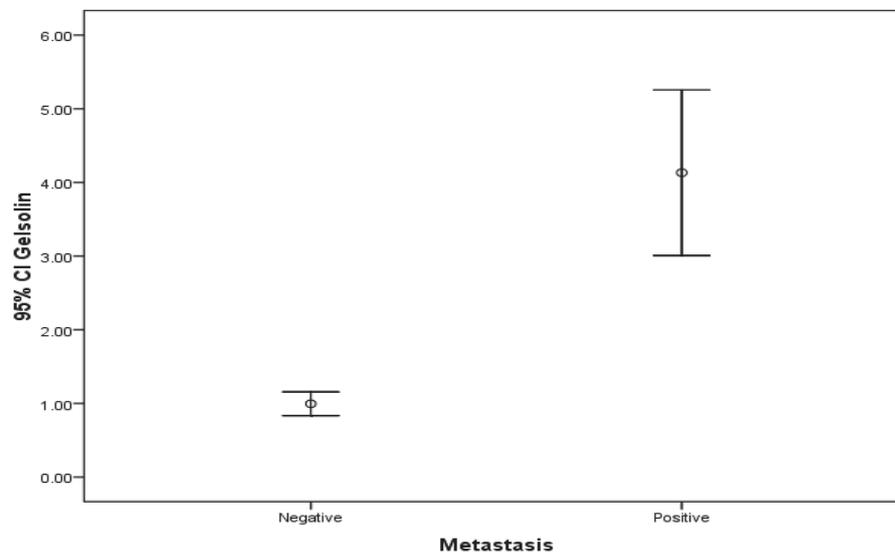
We did not detect a significant correlation between Villin and Gelsolin gene expression levels and patients' age in our study (Table 3).

Table 2. Comparison of Villin and Gelsolin genes expression in both groups

	With metastasis	Without metastasis	P-value
Villin	3.03 \pm 1.35	0.87 \pm 0.88	<0.001
Gelsolin	4.13 \pm 2.4	1 \pm 0.35	<0.001

Table 3. Correlation between Villin and Gelsolin levels and patients' age

	With metastasis	Without metastasis
Villin	R=0.15, P=0.51	R=0.12, P=0.59
Gelsolin	R=0.021, p=0.9	R=0.93, P=0.18

**Fig. 1.** Comparison of Villin gene expression in both groups**Fig. 2.** Comparison of Gelsolin gene expression in both groups

Discussion

The results of this study were indicative of significantly higher levels of Villin and Gelsolin gene expressions in breast cancer patients with axillary lymph node metastasis rather than cases without axillary lymph node metastasis. We also found that their expression level was not significantly correlated with patients' age thus the expression levels of the

mentioned genes were higher in both old and young affected cases.

In a previous study, Baig *et al.* analyzed the expressional profile of Gelsolin gene in tissues obtained from human breasts. They found that the expression level of the Gelsolin gene was down-regulated in breast cancer tissues and was associated

with metastatic growth as well as death in breast cancer cases. In other words, Gelsolin gene expression was lower in the breast cancer group compared to control group and mismatching our findings, the level was even lower in patients with metastatic involvements (8).

In another study, Tutunchi *et al.* assessed Gelsolin gene expression in 70 female breast cancer cases suggesting an increase in Gelsolin gene expression in breast cancer patients. Overexpression of Gelsolin gene is a leading cause of axillary lymph node metastasis in patients with breast cancer. There was not a significant relationship between Gelsolin gene expression levels and tumor size or tumor histologic grades (20).

Another study by Zhu *et al.* investigated the expression levels of five types of peptides including Gelsolin in patients with non-small cell lung cancer (NSCLC). They noticed that the increased Gelsolin gene expression level was associated with lymph node metastasis, malignant disease progression as well as increased risk of mortality (21).

Abedini *et al.* assessed the relationship between Gelsolin gene expression and clinical prognosis as well as survival in women with ovarian serous adenocarcinoma. They reported higher Gelsolin gene expression in women with ovarian cancer which was associated with more tumor growth and poorer clinical prognosis. In other words, they realized that Gelsolin overexpression was notably associated with more invasive tumor behavior and higher cancer mortality rates (22).

The results also suggested significantly higher levels of Villin gene expression in patients with lymph node metastasis than the other group.

In a study by Mohammadpour *et al.*, 42 breast cancer patients' samples were investigated and expression levels of Villin and HER-2 genes were detected with quantitative PCR using pre-designed primers. They realized that Villin gene expression was positively correlated with HER-2 and aggressive tumor nature. Hence, Villin gene overexpression may be a superior predicting element in invasive breast cancer cases (19).

Ma *et al.* studied the relationship between Ezrin protein (Villin) expression and the carcinogenesis as well as prognosis of breast invasive ductal carcinoma. They found marked Ezrin protein (Villin) expression in breast invasive ductal carcinoma which was positively associated with axillary lymph node involvement. They also suggested that high expression levels of Ezrin protein (Villin) may be utilized as a good predicting factor for unfavorable prognosis in the patients with infiltrating ductal breast carcinoma (23).

Yang also conducted a study evaluating Villin gene expression in 52 breast ductal carcinoma patients. Contrary to our results, he found that Villin was absent in all breast cancer cases (24).

Gelsolin and Villin have basic roles in cytoskeleton regulation. Gelsolin is associated with motility, apoptosis, and is indicative of cancer phenotype while it is considered to be an activator and an inhibitor of apoptosis in different cancers (13, 22, 25).

Our results reveal that Gelsolin gene expression is significantly higher in breast cancer cases with axillary lymph node metastasis which highlights greater stages of the disease in these cases. In addition, elevated Gelsolin gene expression level is not correlated with patients' age.

Villin has a crucial role in actin reorganization and cell remodeling so that it affects cancerous cells migration and invasion (26, 27). Villin is an actin-binding protein expressed in several organs of gastrointestinal system such as stomach, pancreas gland, bile ducts and intestines and its higher concentrations are also detected in kidney, lung, endometrium, and ovary adenocarcinomas (28-30).

Our findings highlight that Villin gene expression is significantly higher in axillary lymph node metastatic breast cancer patients in comparison with those without lymphatic involvement, and it is not correlated with patients' age either.

This study had some limitations. Firstly, it was conducted in a tertiary center. Secondly, our sample size was limited. Thirdly, we did not evaluate hormone receptors, HER2 as well as other genes expressions in our study. So larger multi-centric studies along with evaluating the expression of the more genes are recommended. In other words, future investigations will be needed to reach more comprehensive correlations and results.

Conclusion

Villin and Gelsolin genes expression were significantly higher in female breast cancer patients with axillary lymph node metastasis in comparison with breast cancer patients who did not deal with such involvement.

Acknowledgements

We would like to kindly appreciate technicians and administrative staff of the Pathology Department and Iran National Tumor Bank at Imam Khomeini Hospital.

Conflict of Interest

The authors confirm that there are no known conflicts of interest associated with this publication, and there has been no significant financial support for this work that could have influenced its outcome.

References

1. Hassani-pour S, Maghsoudi A, Rezaeian S, Arab-Zozani M, Mokhtari AM, Abdzadeh E, et al. Survival Rate of Breast Cancer in Eastern Mediterranean Region Countries: A

- Systematic Review and Meta-Analysis. *Ann Glob Health*. 2019;85(1):138. [DOI:10.5334/aogh.2521] [PMID] [PMCID]
2. Key TJ, Verkasalo PK, Banks E. Epidemiology of breast cancer. *Lancet Oncol*. 2001;2(3):133-40. [DOI:10.1016/S1470-2045(00)00254-0]
 3. Imanian M, Imanian M, Karimyar M. Sleep Quality and Fatigue among Breast Cancer Patients Undergoing Chemotherapy. *International journal of hematology-oncology and stem cell research*. 2019;13(4):196-200. [DOI:10.18502/ijhoscr.v13i4.1894] [PMID] [PMCID]
 4. Draper L. Breast cancer: trends, risks, treatments, and effects. *AAOHN J*. 2006;54(10):445-51; quiz 52-3. [DOI:10.1177/216507990605401004] [PMID]
 5. Lu C, Sun H, Huang J, Yin S, Hou W, Zhang J, et al. Long-term sleep duration as a risk factor for breast cancer: evidence from a systematic review and dose-response meta-analysis. *Biomed Res Int*. 2017;2017. [DOI:10.1155/2017/4845059] [PMID] [PMCID]
 6. Sadoughi F, Lotfnezhad Afshar H, Olfatbakhsh A, Mehrdad N. Application of Canonical Correlation Analysis for Detecting Risk Factors Leading to Recurrence of Breast Cancer. *Iran Red Crescent Med J*. 2016;18(3):e23131. [DOI:10.5812/ircmj.23131]
 7. Tao Z, Shi A, Lu C, Song T, Zhang Z, Zhao J. Breast Cancer: Epidemiology and Etiology. *Cell Biochem Biophys*. 2015;72(2):333-8. [DOI:10.1007/s12013-014-0459-6] [PMID]
 8. Schneeweiss A, Lux MP, Janni W, Hartkopf AD, Nabieva N, Taran F-A, et al. Update breast cancer 2018 (part 2)-advanced breast cancer, quality of life and prevention. *Geburtshilfe Frauenheilkd*. 2018;78(3):246. [DOI:10.1055/s-0044-101614] [PMID] [PMCID]
 9. Group EBCTC. Effect of radiotherapy after breast-conserving surgery on 10-year recurrence and 15-year breast cancer death: meta-analysis of individual patient data for 10 801 women in 17 randomised trials. *The Lancet*. 2011;378(9804):1707-16. [DOI:10.1016/S0140-6736(11)61629-2]
 10. Madison BB, Dunbar L, Qiao XT, Braunstein K, Braunstein E, Gumucio DL. Cis elements of the villin gene control expression in restricted domains of the vertical (crypt) and horizontal (duodenum, cecum) axes of the intestine. *J Biol Chem*. 2002;277(36):33275-83. [DOI:10.1074/jbc.M204935200] [PMID]
 11. Yang Z. The utility of villin and mammaglobin in the differential diagnosis between intrahepatic cholangiocarcinoma and breast cancer. *Applied immunohistochemistry & molecular morphology*. 2015;23(1):19-25. [DOI:10.1097/PAI.0000000000000035] [PMID]
 12. Hoskin V, Szeto A, Ghaffari A, Greer PA, Cote GP, Elliott BE. Ezrin regulates focal adhesion and invadopodia dynamics by altering calpain activity to promote breast cancer cell invasion. *Mol Biol Cell*. 2015;26(19):3464-79. [DOI:10.1091/mbc.E14-12-1584] [PMID] [PMCID]
 13. Baig RM, Mahjabeen I, Sabir M, Masood N, Ali K, Malik FA, et al. Mutational spectrum of Gelsolin and its down regulation is associated with breast cancer. *Dis Markers*. 2013;34(2):71-80. [DOI:10.1155/2013/795410] [PMID] [PMCID]
 14. Winston JS, Asch HL, Zhang PJ, Edge SB, Hyland A, Asch BB. Downregulation of gelsolin correlates with the progression to breast carcinoma. *Breast Cancer Res Treat*. 2001;65(1):11-21. [DOI:10.1023/A:1006446108411] [PMID]
 15. Asch HL, Winston JS, Edge SB, Stomper PC, Asch BB. Down-regulation of gelsolin expression in human breast ductal carcinoma in situ with and without invasion. *Breast cancer research and treatment*. 1999;55(2):177-86. [DOI:10.1023/A:1006203632228] [PMID]
 16. Asch HL, Winston JS, Edge SB, Stomper PC, Asch BB. Down-regulation of gelsolin expression in human breast ductal carcinoma in situ with and without invasion. *Breast Cancer Res Treat*. 1999;55(2):177-86. [DOI:10.1023/A:1006203632228] [PMID]
 17. Noske A, Denkert C, Schober H, Sers C, Zhumabayeva B, Weichert W, et al. Loss of Gelsolin expression in human ovarian carcinomas. *Eur J Cancer*. 2005;41(3):461-9. [DOI:10.1016/j.ejca.2004.10.025] [PMID]
 18. Bansal C, Singh US, Misra S, Sharma KL, Tiwari V, Srivastava AN. Comparative evaluation of the modified Scarff-Bloom-Richardson grading system on breast carcinoma aspirates and histopathology. *CytoJournal*. 2012;9:4. [DOI:10.4103/1742-6413.92550] [PMID] [PMCID]
 19. Mohammadpour H, Hashemi M, Saffari M, Razavi ANE. Comparative Analyses of Villin and HER-2 Genes Expression in Breast Cancer. *Archives of Breast Cancer*. 2015:120-4.
 20. Tutunchi S, Saffari M, Mazaheri M, Nourmohammadi P, Memari F, Shirkoobi R, et al. Evaluating Gelsolin Gene Expression Among Iranian Breast Cancer Patients. *Bas & ClinCancer Res*. 2018;10(4):16-25.
 21. Zhu WY, Hunag YY, Liu XG, He JY, Chen DD, Zeng F, et al. Prognostic evaluation of CapG, gelsolin, P-gp, GSTP1, and Topo-II proteins in non-small cell lung cancer. *Anat Rec (Hoboken)*. 2012;295(2):208-14. [DOI:10.1002/ar.21523] [PMID]
 22. Abedini MR, Wang PW, Huang YF, Cao M, Chou CY, Shieh DB, et al. Cell fate regulation by gelsolin in human gynecologic cancers. *Proc Natl Acad Sci U S A*. 2014;111(40):14442-7. [DOI:10.1073/pnas.1401166111] [PMID] [PMCID]
 23. Ma L, Zhang XH, Xing LX, Li YH, Wang XL, Wang YJ. [Relationship of ezrin protein expression to the carcinogenesis and prognosis of infiltrating breast ductal carcinoma]. *Zhonghua Zhong Liu Za Zhi*. 2008;30(4):279-83.
 24. Yang Z. The utility of villin and mammaglobin in the differential diagnosis between intrahepatic cholangiocarcinoma and breast cancer. *Appl Immunohistochem Mol*. 2015;23(1):19-25. [DOI:10.1097/PAI.0000000000000035] [PMID]
 25. Deng R, Hao J, Han W, Ni Y, Huang X, Hu Q. Gelsolin regulates proliferation, apoptosis, migration and invasion in

- human oral carcinoma cells. *Oncol Lett.* 2015;9(5):2129-34. [[DOI:10.3892/ol.2015.3002](https://doi.org/10.3892/ol.2015.3002)] [[PMID](#)] [[PMCID](#)]
26. Omran OM, Al Sheeha M. Cytoskeletal Focal Adhesion Proteins Fascin-1 and Paxillin Are Predictors of Malignant Progression and Poor Prognosis in Human Breast Cancer. *J Environ Pathol Toxicol Oncol.* 2015;34(3):201-12. [[DOI:10.1615/JEnvironPatholToxicolOncol.2015013663](https://doi.org/10.1615/JEnvironPatholToxicolOncol.2015013663)] [[PMID](#)]
27. Arango D, Al-Obaidi S, Williams DS, Dopeso H, Mazzolini R, Corner G, et al. Villin expression is frequently lost in poorly differentiated colon cancer. *Am J Pathol.* 2012;180(4):1509-21. [[DOI:10.1016/j.ajpath.2012.01.006](https://doi.org/10.1016/j.ajpath.2012.01.006)] [[PMID](#)] [[PMCID](#)]
28. Moll R, Robine S, Dudouet B, Louvard D. Villin: a cytoskeletal protein and a differentiation marker expressed in some human adenocarcinomas. *Virchows Archiv B.* 1987;54(1):155-69. [[DOI:10.1007/BF02899208](https://doi.org/10.1007/BF02899208)] [[PMID](#)]
29. Batra S, Vaid AK, Bhargava R, Singh A. Malignancy knows no boundaries. *Case Reports.* 2013;2013:bcr2013010383. [[DOI:10.1136/bcr-2013-010383](https://doi.org/10.1136/bcr-2013-010383)] [[PMID](#)] [[PMCID](#)]
30. Bacchi CE, Gown AM. Distribution and pattern of expression of villin, a gastrointestinal-associated cytoskeletal protein, in human carcinomas: a study employing paraffin-embedded tissue. *Lab Invest.* 1991;64(3):418-24.

How to Cite This Article

Borhan, A. et al. Evaluation of the Relationship Between Expression of Villin and Gelsolin Genes and Axillary Lymph Node Metastasis in Patients with Breast Cancer. *Iranian Journal of Pathology*, 2021; 16(1): 00-00. doi: