# **Original Article**

# Microbial Etiology and Antimicrobial Susceptibility of Bactria Implicated in Urinary Tract Infection in Tehran, Iran

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#### ABSTRACT

Background and Objectives: Urinary tract infections (UTI) are one of the most common infectious diseases with different microbial agent and antimicrobial resistant pattern in hospitalized patients and outpatients. In order to assess the adequacy of therapy, knowledge of prevalence and resistance pattern of the bacteria is necessary. The main aim of this study was to evaluate the prevalence and the antimicrobial resistance pattern of main bacterial responsible for UTI in order to establish an appropriate empirical therapy.

*Methods:* All urine samples were referred to Imam Hospital Laboratory, Tehran, Iran during 2011-2012, urine culture isolated and bacteria were identified and the profile of antibiotic susceptibility was characterized.

Result: From 1851 urine cultures, UTI was more frequent in woman (68%) E. coli was as usual the most common pathogen implicated in UTI. Most susceptibility was to imipenem (98.9%). nitroforantoin (96%) and amikacin (94.1%) and increased resistance to penicillin (66.6%), nalidixic acid (62.1%) ampicilin (60.1%) and cotrimoxazole 54.3%.

Discussion: The most common isolated pathogen was E. coli. According to antibiogram susceptibility, the recommended antimicrobial drugs are nitroforantoin and imipenem. nalidixic acid and cotrimoxazole are not recommended because drug resistance is high.

Keywords: Antibiogram, Urinary Tract Infection, Iran

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Trinary tract infection (UTI) is one of the most common infectious diseases of community and also of the hospital setting resulting in high rates of morbidity. Fifty percent of women experience urinary tract infection at least once in their lifetime (1). Some risk factors for UTI are female sex, elevated age, pregnancy and diabetes (1-3).

The most common infection in hospital is UTI due to catherization. Besides, antimicrobial misuse in clinical medicine has increased the microbial resistance and consequently spread bacterial resistance strains. That is a serious public health problem (3-5).

The main cause of UTI is uropathogens such as *E*. coli (46.4%-74.2%), Klebsiella SPP. (6-13.45%) Proteous SPP. (4.7-11.9%) and Entrococcous SPP. (5.3-9.54%). E. coli has been identified as the most frequent pathogen in the uncomplicated patient. But Proteous, Klebsiella, Entrobacter, Seratia and Pseudomonas isolated in recumbent, complicated and catheterized patient. The early treatment of UTI decreased the rate of morbidity. (1, 2, 6-9). In order to prescribe the appropriate antibiotics and prevent antibiotic resistance in patients, it would be necessary to know the model of frequency of microorganisms causing urine infections as well as the model of antibiotic sensitivity and resistance of the microorganisms. Unfortunately there is little publication about the main uropathogen in community acquired UTI and antimicrobial resistance pattern when compared with UTI at hospital level.

Given the growing drug resistance in microorganisms, once-effective antibiotics are less effective on the bacteria causing urinary tract infection. It results from the emergence and expansion of bacteria-resistant strains due to the genetic properties of bacteria, population growth, travel and non-standard administration of antibiotics. The report of sensitivity to anti-microbial factors is submitted to the doctor most often 48 hours after the sample is delivered to the lab. Therefore, in most cases, the

treatment is done based on experience and since experimental antibiotic treatment of urinary tract infections must be based on epidemiology and the resistance pattern of uropathlogen, this study is necessary to be conducted.

This information is very important and implies a periodic monitorization in order to decrease the number of therapeutic failure (6-8). The main aim of this study was to evaluate the prevalence and the antimicrobial resistance pattern of the main bacteria responsible for UTI in order to establish an appropriate empirical therapy.

#### **Materials and Methods**

We evaluate urine cultures of inpatients & outpatients referred to clinical laboratory of Imam Hospital, Tehran, Iran during the period of 2012-2013. For each patient, the collection date, age, sex, related ward, culture result, identification of bacterial strain and antimicrobial susceptibility test was done. The method of sampling for culture and antibiogram in patients with the urinary control was taking mid-stream sample and from children without sphincter controlling the suprapubic aspiration were done. All samples were studied by a veteran expert in the lab and with the application of standard methods.

The urine sample was inoculated in different culture media. A calibrate loop was dripped in vertical position in the urine sample and the loop was used to inoculate the plate. After incubation, the urine culture when monomorplic bacteria growth was higher than 105 cfu/ml, the culture was classified as positive. Additional biochemical tests were performed on the morphology of the isolated bacterial on the results of the microscopic examinations of the gram stained smear. A bacterial suspension in physiology saline solution with turbidity at 0/5 on McFarland scale was prepared. The suspension was spread with a swab on Muller-Hinton agar and antimicrobial disk were placed onto to medium according to isolated organism. The plates were incubated at 37 °C for 18-24 h, after incubation the antimicrobial

efficacy was determined by measuring of diameter of the zones of inhibition.

#### Isolation and Identification of Bacteria

Isolation of these bacteria was performed using Streak-Plate Method on Blood agar and MacConkey agar culture with standard loop (internal diameter 34/1 mm). Culture plates were incubated at 37°C for 24 h. The cultures consisting of more than 10<sup>5</sup> colonies of particular bacteria were considered as positive cultures. The incubation of negative 24 h cultures was extended for another 24 h. Bacteria were identified through performing biochemical tests (indole, citrate, oxidase, and production of H<sub>2</sub>S, lysine decarboxylase, fermentation of lactose, urea hydrolysis, gas production, catalase, coagulase, mannitol fermentation and susceptibility testing novobiocin).

### **Antibiotic Susceptibility Testing**

Strains drug resistance evaluation was carried out using disk diffusion method on Mueller-Hinton medium (Merck, Germany). After inoculating the bacteria on Muller-Hinton agar and placing the antibiotic disks, plates were incubated for 24 h in incubator. Then, according to the size of the growth inhibition zone around the disks and international numbers of (NCCLS), results were categorized and reported in three groups :susceptible and Sensitive (S), intermediate susceptibility or sensitivity (I) and resistance (R). E. coli ATCC 25922, Pseudomonas aeruginosa ATCC 27853 and Staphylococcus aureus ATCC 25923 were used as control strains and the test results were only accepted when the inhibition zone diameters of the above mentioned control strains were within performance ranges (as described by CLSI No:M100-S16). In case of mixed bacteria, only the major and predominant pathogens were tested. The antibiotics used for susceptibility testing were norfloxacin 10 µg, ofloxacin 5 µg, ciprofloxacin 5 µg, nitrofurantoin 300 µg, co-trimoxazole (SXT), carbenicillin 100 μg, ampicillin 10 μg, cephalothin30 μg, gentamicin 10  $\mu$ g, amikacin 30  $\mu$ g, nalidixic acid 30  $\mu$ g, cefotaxime 30  $\mu$ g, imipenem 10  $\mu$ g, tetracycline 30  $\mu$ g, penicillin 10 IU, oxacillin 1  $\mu$ g, vancomycin 30  $\mu$ g, ceftriaxone 30  $\mu$ g, and ticarcillin75  $\mu$ g.

This study has been endorsed by the Ethics Committee of the hospital and the university. Written informed consent was obtained from all patients and the research and ethic committee of TUMS approved the study protocol on human subject.

Statistical Package for Social Sciences were used to analyze data of the current study with employment of student t-test for quantitative and Chi square test for qualitative variables while the values were considered statistically significant at a P<0.05.

#### Result

From the 1851 samples, 1257 (68%) were female, also 1666 (90%) were more than 12 year old and 1041 (56.2%) related to hospitalized patients (Table 1).

**Table 1-**Age & Sex distribution of patients

Age (yr)	Male (594)	Female (1257)				
<12	70 (37.8%)	115 (62.2%)				
>12	524 (31.5%)	1142 (68.5%)				

The main agents of UTI were isolated from outpatients were *E. coli* (51.5%), *S. hemolyticus* and *S. aureus*, and from inpatient, *E. coli* (58%), *Candida* and *Entrobacter* were most common organism (Table 2).

The most common isolated organism in patients below 12 year were *E. coli*, *Entrobacter*, coagulase negative, *Staphylocuccus* and *Proteous* and in above 12 years were *E. coli*, *Staphylocuccu* coagulase negative and *Candida*. Similarly, *E. coli*, *Staphylocuccu* coagulase negative and *Candida* were predominant agents in both sexes. In hospitalized patients, *E. coli* was predominant

in Renal Transplant (44.4%), Pediatric (43.3%), Dermatology (51.9%), Urology (41%), Cardiac (53.8%), Emergency (52.2, Internal (43.2%), Surgery (45.7%), Infections (46.3%), Neonate (78.1%) and Gynecology (47.4%) wards. In ICU, the predominant agent was *Candida*, in

BMT section was *Acintobacter* and in Neurology section was *Entrococus*.

The antimicrobial resistance pattern of main bacteria is showed in Table 3. Table 4 shows different antimicrobial resistance of *E. coli* in hospitalized patients and outpatients.

**Table 2-** Distribution of isolated bacteria from inpatients & outpatients

Bactria	Out patient (isolated Bactria %)	Hospitalized Patient (Isolated Bactria %)		
E. coli	51.5	58		
Candida	3	12		
Entrobacter	2	8		
Staphylococcus hemolyticus	0.5	5		
Staphylococcus aureus	3	5		
Others	33	21		

**Table 3-** The antimicrobial resistance pattern of main bacteria

Microbial Agent Antibiotic	E. coli (%)	Klebsiella(%)	Entrobacter(%)	Entococuss(%)	Pseudomonas(%)	Acintobacter(%)	Streptococcus nonhemolyticus(%)	Protcous(%)	Staphylococus Epidermidis(%)	Staphylococus hemolyticus(%)	Staphylococus saprophyticus(%)	Staphylococus areous(%)	Citrobacter(%)
Imipenem	sen	-	-	-	Sen*	-	-	Sen	Res**	-	-	Res	-
	98				95			94	100			66.6	
Nitroforantoin	Sen	-	Sen	-	Res	-	-	Sen	-	Sen	-	-	-
	95.5		82.3		83.4			83.3		9.6			
Amikacin	Sen	Sen	Sen	-	Sen	-	-	-	Sen 96	-	-	-	Sen
	94.1	92	86.7		88								83.3
Nalidixic acid	Res	-	-	-	-	-	-	Res	-	-	-	-	-
	62.1							66.6					
Gentamycin	-	Sen	-	-	Sen	Sen	Res	-	-	-	-	Sen	-
		92.5			81.4	94.4	55					99.8	
Vancomycin	-	-	-	-	-	-	-	-	Sen 98	-	-	-	-
Ampicilin	Res 60	Res	Res	-	-	-	Sen	Res	-	-	-	-	Res
		72	69				90	66.6					60
Ceftazidim	Res 87	Res	Res	-	Res	-	Res	Res	Res 72	Res	-	Res	Res
		77	96		90		60	75		100		75	100
Ceftriaxon	-	-	-	-	-	-	-	-	Sen 94	Res 71	-	-	-
Penicilin	Res	-	-	-	-	-	-	-	-	-	-	-	-
	66.6				_							_	
Co- trimoxazde	-	-	-	-	Res			-	-	-	-	Res	-
					95							48.9	
Erythromycin -	-	-	-	-	-	-	-	-	-	-	Res	-	-
											100		
Ciprofluoxacin	-	-	-	-	-	-	-	-	-	-	sen	-	-

<sup>\*:</sup> Sensitive

<sup>\*\*:</sup> Resistant

E. coli Resistance to Outpatient (%) Hospitalized patient (%) **Ampicilin** 50.4 81.6 26.7 Gentamycin 15.7 Nalidixic acid 40 76.8 Nitroforantoin 3.7 6.8 6.9 Amikacin 4.7 Ceftriaxon 37.7 71.4 Ciprofluoxacin 46.9 30.8 Ceftazidim 91.2 30.6 Co-trimoxazole 47.4 67.6 **Imipenem** 0.5 2.2

**Table 4-** Different *E. coli* antimicrobial resistance in hospitalized and out patients

#### Discussion

This study evaluated species distribution and antibiotic susceptibility of urinary tract infection (UTI) isolated in Imam Hospital of Tehran, Iran. *E. coli* was the most frequent uropathogen (being implicated in more than one half of all the UTI). Similar frequency of isolates of *E. coli* has been obtained in studies performed in Latin American (52%), Norway (56.7%), great Britain (65.1%) and USA (68%) (1, 9-14).

Although E. coli was the most common uropathogen in both sexes, its incidence was significantly higher in woman (P=0.0006). The question of an underlying urinary pathology of infection in female as compared to male cases was beyond the scope of this study.

Pseudomonas aeruginosa is most frequent in male due to particular characteristic inherent to the patient including sex, use of antimicrobial agent, previous intervention in urinary tract and patient with neurogenic bladder, but in this study uropathogens were similar in both sexes (1, 3, 14-16).

In this study it was not observed significant difference among the bacteria responsible for these infections in the different age groups.

In outpatients, after *E. coli*, *S. hemolyticus* and *S. areous* were common uropathogen but in hospitalized patients, *Entrobacter* and *Candida* was common pathogen due to more invasive

intervention on urinary tract. This finding correlates with previous studies in Norway and Iran (2, 13).

In most wards of the hospital, *E. coli* was most common isolated uropathogen but in some ward other species were isolated.

Although *E. coli* was responsible for more than half of the UTI, its antimicrobial resistance was significantly lower than that presented by the other bacteria (1, 2, 16). In this study, E. coli had the most resistance to penicillin, ampicilin, nalidixic acid and Co-trimoxazole but most sensitivity to imipenem, nitroforantoin and amikacin is reported. This finding is compatible with study of Keah with high resistant E. coli to ampicilin (63%) and cotrimoxazole (43%) (6).-Also Peterson J study shows resistance to ampicilin (50.1%) and cotrimoxazole (22.1%) (10). The result antimicrobial resistance indicate that cotrimoxazole, nalidixic acid and ampicilin should be ignored to treat of UTI as the resistance rate was higher than the recommended value (<20%) indicated (1, 2, 17-20).

The drugs, once effective in treatment of patients with urinary tract infection, have now become resistant and their effectiveness has declined. Comparison our findings with those from other studies showed that the drug resistance of bacteria causing urinary tract infections in this study is much higher than in most parts of the world

and the drugs prescribed for these infections are ineffective.

While these drugs can still be effective in some parts of the world, this study is recommended to be conducted in every region and every several years so that more appropriate decisions could be made for the treatment of these patients before antibiogram testing.

Besides, when antibiotic resistance is studied, such factors as anomalies in the urinary system, urinary reflux, recent administration of antibiotics and recurrent urinary infection must be taken into account because some of these factors are likely to cause serious problems.

As expected, hospitalized patients have high rate of resistance due to more Antibiotic consumption and more unusual organism due to invasive intervention compared with out patients and attention to antibiogram resistance pattern could be helpful (15,16).

#### Conclusion

The most common isolated pathogen was *E. coli*. According to antibiogram susceptibility, the recommended antimicrobial drugs are nitroforantoin and imipenem. nalidixic acid and cotrimoxazole are not recommended because drug resistance is high. Antibiotic resistance in this study was remarkable and it must be taken into account. Any unnecessary prescription of antibiotics for patients must be avoided.

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#### References

1. Linhares *et al.* frequency and antimicrobial resistance pattern of bacterial implicated in community urineary

tract infections, a ten year surveillance study 2000-2009. BMC Infections Disease J 2013;13(19).

- 2. Saderi H, Owlia P, Jalali Nadushan MR, Zaeri F, Zandieh E. A tree-year study of demographic characteristics of patients with urineary tract infection, microbial etiology, and susceptibility of isolated bacteria to antibiotics in shaheed Mostafa Khomeini hospital. Iranian J Pathol 2006; 1(3):99-104.
- 3. Orrett FA, Davis GK. A comparison of antimicrobial susceptibility profile of urineary pathogens for the years, 1999 and 2003. West Indian Med J 2006; 55(2):95-9.
- 4. Hasan AS, Nair D, Kaur J, Baweja G, Deb M, Aggarwal P. Resistance patterns of urineary isolates in a tertiary Indian hospital. J Ayub Med Coll Abbottabad 2007; 19(1):39-41.
- 5. Walter E. Stamm.Urineary tract infection. Kasper D, Braunwald E, Fauci A, Hauser S, Longo D, Jameson J.L. In: Harrison's priciples of internal medicine. 16<sup>th</sup> ed. New York: McGraw-Hill; 2005.
- 6. Keah SH, Wee EC, Chng KS, Keah KC. Antimicrobial susceptibility of community-acquired uropathogens in general practice. Malaisian Family Phisician 2007; 2(2):64-9.
- 7. De Francesco MA , Ravizzola G , Peroni L , Negrini R , Manca N .Urineary tract infections in Brescia,Italy:Etiology of uropathogens and antimicrobial resistance of common uropathogens. Med Sci Monit 2007;13(6):BR136-144.
- 8. Akram M, Shahid M, Khan AU. Etiology and antibiotic resistance patterns of community-acquired urineary tract infection in JNMC Hospital Aligarh, India. Ann Clin Microbiol Antimicrob 2007; 6:4.
- 9. Kamat US, Ferreira AMA, Savio R, Motghare DD. Antimicrobial resistance among nosocomial isolates in a teaching hospital in Goa. Indian J Community Med 2007; 33(2):89-92
- 10. Peterson J, Kaul S, Khashab M, Fisher A, Kahn JB. Identification and pretherapy susceptibility of pathogens in patients with complicated urineary tract infection or acute pyelonephritis enrolled in a clinical study in the United States from November 2004 through April 2006. Clin Ther 2007; 29(10):2215-21.

- 11. De Backer D, Christiaens T, Heytens S, De Sutter A, Stobberingh EE, Verschraegen G. Evolution of bacterial susceptibility pattern of *Escherichia coli* in uncomplicated urineary tract infection in a country with high antibiotic consumption: a comparison of two surveys with a 10 years interval. J Antimicrob Chemother 2008; 62:364-8.
- 12. Randrianirina F, Soares JL, Carod JF, Ratsima E, Thonnier V, Combe P, *et al.* Antimicrobial resistance among uropathogens that cause community-acquired urineary tract infections in Antananarivo. Madagascar.J Antimicrob Chemother 2007;59:309-12.
- 13. Grude N.Tveten Y, Kristiansen BE.Urineary tract infection in Norway, Bacterial etiology and susceptibility. A retrospective study of clinical isolates. Clinic Microbial infect 2001 7:543-7.
- 14. Zhanel GG, Hisanaga TL, Laing NM, DeCorby MR, Nichol KA, Palatnick LP, *et al.* Antibiotic resistance in outpatiant urineary isolates:final results from the North American University Infection Collaborative Alliance(NAUTICA). Int J Antimicrob Agents 2005; 26:380-8.
- 15. Pires MC, Frota Kde S, Martins Junior Pde O, Correia AF, Cortez-Escalante JJ, Silveria CA. Prevalence and bacterial susceptibility of community acquired urineary tract infection in university hospital of Brasilia, 2001 to

- 2005. Rev Soc Bras Med Trop. 2007;40(6):643-7.
- 16. Leblebicioglu H, Esen S; Turkish Nosocomial Urineary Tract Infection Study Group. Hospital-acquired urinary tract infection in Turkey: a nationwide multicenter point prevalence study. J Hosp Infect 2003; 53:207-10.
- 17. Dias Neto JA, Magalhaes da Silva LD, Pereira Martins AC, Tiraboschi RB, Alonso Domingo AL, Suaid HJ, *et al.* Prevalence and bacterial susceptibility of hospital acquired urinary tract infection. Acta Cirurgica Brasileira 2003;18(supl.5):36-8.
- 18. Jose Anastacio Dias Neto JA, Martins AC, Magalhaes da Silva LD, Tiraboschi RB, Alonso Domingos AL, Cologna AJ, *et al.* Community acquired urineary tract infection: etiology and bacterial susceptibility. Acta Cirurgica Brasileira 2003; 18(supl. 5):33-6.
- 19. Andrade SS, Sader HS, Jones RN, Pereira AS, Pignatari AC, Gales AC. Increased resistance to first-line agents among bacterial pathogens isolated from urineary tract infections in Latin America:time for local guidelines! Mem Inst Oswaldo Cruz,Rio de janeiro 2006;101(7):741-8.
- 20. Colodner R, Keness Y, Chazan B, Raz R. Antimicrobial susceptibility of community-acquired uropathogens in northern USA. Intern J Antimicrob Agents 2001; 18(2):189-92.